Inhibition of Germination and Seedling Growth of Rice by Culture Filtrate of Aflatoxigenic Aspergillus flavus

Shubhransu Nayak¹, Urmila Dhua¹, Chandan Sengupta², Soma Samanta¹ and Sudhi Ranjan Dhua¹

 Crop Protection Division, Central Rice Research Institute, Cuttack, Odisha, India-753006, Email: udhua.crri@gmail.com; (2) Microbiology & Virology Section, Department of Botany, University of Kalyani, Kalyani, Nadia, West Bengal, India, 741235

Abstract

Nayak, S., U. Dhua, C. Sengupta, S. Samanta and S.R. Dhua. 2015. Inhibition of germination and seedling growth of rice by culture filtrate of aflatoxigenic *Aspergillus flavus*. Arab Journal of Plant Protection, 33(1): 93-95.

Aflatoxin B1 produced by *Aspergillus flavus* causes damage to crops and cereals including rice by inhibiting seed germination, elongation of the hypocotyls or roots and alters many physiological processes in developing seedlings. Rice cultivar Savitri is a popular high yielding variety in the eastern coastal part of India, where favorable growth condition for *A. flavus* exist and the harvesting practice of the farmers usually result in the infection of rice crops with this fungus. Hence, the effect of culture filtrate of *A flavus* on cv. Savitri (CR 210-1009) was investigated. Seed germination, root shoot length and root shoot weight of Savitri seedlings were drastically reduced by dipping the seeds in the undiluted and diluted (50% and 25%) culture filtrate. In the case of undiluted culture filtrate, seed germination rate and seedlings mortality rate were 18% and 75% compared to 70% and 0.00% for the untreated control, respectively. The residual seed weight was lower (358.9mg) in the case of untreated control and higher in case of undiluted filtrate (521mg). Results indicated that the culture filtrate containing the aflatoxin, interfered with the physiological processes which prevented the use of endosperm by the developing seedlings.

Keywords: Aflatoxins, Savitri, germination, Aspergillus flavus, rice.

Introduction

Aflatoxins are biologically active secondary metabolites produced mainly by A. flavus and have been detected in cereal grains, oil seeds, fermented beverages made from grains, milk, cheese, meat, nut products, fruit juice and numerous other agricultural commodities (1). Though mycotoxins and particularly aflatoxin contamination is less commonly reported for rice than for many other cereal crops, rice represents a very good substrate for fungal growth and toxinogenesis since it is used as an ideal culture medium to test the toxigenic potential of isolated strains. The major mycotoxigenic fungi in rice are Aspergillus spp., Fusarium spp. and Penicillium spp. (2). Besides its toxigenic effect on animals, Aflatoxin B1 inhibits seed germination, seedling growth and alters the physiological processes of seedlings of many crops including cereals such as rice (3, 4) wheat (5) and maize (1). Aflatoxins may inhibit seed germination, elongation of the hypocotyls or roots of developing seedlings, or both, and by interference with chlorophyll synthesis in certain plants (6). The harmful effects of such fungal invasion may be glume or grain discoloration, loss in viability, quality and toxin contamination (1).

The aflatoxin producing fungi are widely distributed in nature and can grow over a wide range of environmental conditions (7). However, high relative humidity and temperature favors the growth and aflatoxin production (8, 9). In the eastern coastal part of India where such type of environmental conditions exist, farmers usually keep harvested paddy in the field for 4-5 days for proper sun drying. The occurrence of sudden heavy rain during the harvesting season resulting in flood like situation, makes the rice crop vulnerable to infection with *Aspergillus* spp. (10) and favors the growth and spread of *Aspergillus flavus* and subsequent aflatoxin contamination of the rice crop. Among the various stages, rice at the drying stage and the stage preceding milling were shown to contain aflatoxins (11). Accordingly, the effect of dipping seeds in culture filtrate of aflatoxigenic *A. flavus* on seed germination and seedling growth of rice cultivar Savitri (CR 1009, released by Central Rice Research Institute, India) was investigated.

Materials and Methods

Aspergillus flavus isolate A129 was grown in -Cyclodextrine Potato Dextrose Broth (12) for 15 days in 150 ml conical flask at ambient temperature. The mycelial mat was separated and the broth was filtered to be used in the study. Three concentrations of this filtrate such as undiluted culture filtrate (100%) and two dilutions (50% and 25%) of culture filtrate were used. Hundred seeds of rice cv. Savitri was properly dipped in 3 ml of each solution and then plated on blotting paper in plastic petri plates. To each type of plate, 6 ml more solution was added and then kept at ambient temperature. Seed germination and seedling mortality rates, root-shoot length, root-shoot weight and residual seed weight (i.e. the weight of the seeds after removing root and shoot) were measured. The seedling mortality rate was calculated using the formula:

Seed mortality (%) = (Number of dead seeds after germination/Total number of germinated seeds) $\times 100$

Results and Discussion

The A. *flavus* isolate (A129) used in the current study was isolated from a raw dehisced rice sample. It was non-sclerotic type and produced 25 μ g/ml of aflatoxin B1 in culture broth.

The toxic culture filtrate of A129 drastically reduced the seed germination rate, root shoot length and root shoot weight of rice cultivar Savitri and the reduction of these parameters were directly proportional to the concentration of the culture filtrate (Table 1 & Figure 1). The germination rate of seeds soaked in sterilized water (control) was 70%, with no seedlings mortality, whereas in the case of undiluted culture filtrate, germination rate was 18% and seedling mortality rate was 75%. Root growth was completely inhibited in seedlings treated with undiluted filtrate and very poorly developed in 50% and 25% filtrates. Root weight of seedlings treated with undiluted and 50% filtrate were 9 and 13 mg, whereas shoot weight was 133 and 161 mg, respectively (Table 1). These results were in accordance with those obtained previously (1). Aflatoxins in food grains interfere with protein synthesis by inhibiting the incorporation of amino acids into protein, resulting in non-germination of embryo and it also binds to DNA and thus prevents RNA synthesis (13). In some legumes, this reduction in root and shoot length was thought to be due to allantoinase (allantoina amido hydrolase) activity on germinating seeds (14). The residual seed weight was lowest (358.9 mg) in the case of untreated control and it increased with the increase in concentration of culture filtrate, i.e. highest in case of undiluted filtrate (521 mg). It indicated that the aflatoxin interfered with the physiological processes which prevented the use of endosperm by the developing seedlings.

Rice cv. Savitri (CR 210-1009) is a popular high yielding cultivar in the eastern coast of India, grown in shallow low-land ecosystem. It is resistant to major rice diseases and pests and of 145 days growth duration, hence largely adopted for cultivation by farmers. The current study showed that this variety was susceptible to aflatoxin B1 produced by *A. flavus*. Hence, control measures should be adopted to prevent the contamination of this fungus to rice otherwise it may cause quantitative and qualitative changes in chemical composition (biodeterioration) of the rice seeds.

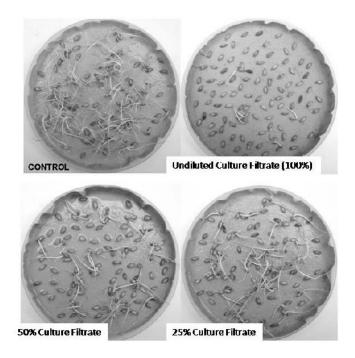


Figure 1. Plates showing germination and mortality of seedlings treated with aflatoxic culture filtrate of *A. flavus*.

Acknowledgement

Authors acknowledge the assistance provided by the Director of Central Rice Research Institute, Cuttack, India for providing all necessary facilities.

Table 1. Effect of cultural extract of toxigenic Aspergillus flavus isolate A129 on germination and mortality rates of rice cultivar Savitri (CR 1009).

Solution	Germination rate (%)	*Seedling mortality rate (%)	Root length (cm)	Shoot length (cm)	Root weight (mg)	Shoot weight (mg)	Residual seed weight (mg)
Control	70 ± 2.9	0.0	4.2 ± 0.19	4.3 ± 0.10	27 ± 1.2	208 ± 11.84	358.9
Cultural filtrate 100%	18 ± 1.4	75 ± 6.88	0.0	1.8 ± 0.13	9 ± 0.46	133 ± 8.24	521.0
Cultural filtrate 50%	32 ± 0.6	22 ± 2.66	0.2 ± 0.05	2.5 ± 0.12	13 ± 1.47	161 ± 1.65	495.0
Cultural filtrate 25%	35 ± 1.2	19 ± 1.45	2.3 ± 0.18	3.6 ± 0.14	22 ± 0.41	205 ± 1.45	468.0

الملخص

ناياك، اس.، ي. دهوا، س. سنكيبتا، أس.. سامان توس و أس.ر. دهوا. 2015. تثبيط إنبات ونمو بادرات الرز برشاحة الفطر Aspergillus flavus المنتج للأفلاتوكسين. مجلة وقاية النبات العربية، 33(1): 93–95.

يسبب الأفلاتوكسين B1 الذي ينتجه الفطر Aspergillus flavus ضرراً للمحاصيل ومحاصيل الحبوب بما في ذلك الرز عن طريق منع إنبات البذور، واستطالة السويقة الجنينية أو الجذور وتغيير العمليات الفيزيولوجية في البادرات المتطورة. يعد صنف الرز سافيتري من الأصناف الشعبية عالية الغلة في الجزء الشرقي الساحلي من الهند، حيث تتوافر ظروف ملائمة لنمو الفطر Aspergillus flavus وتؤدي ممارسات الحصاد التي يتبعها المزارعون عادة إلى الإصابة بالفطر. أظهرت دراسة رشاحة مزرعة الفطر قروف ملائمة لنمو الفطر CR 210-1009 وتؤدي ممارسات الحصاد التي يتبعها المزارعون عادة إلى الإصابة بالفطر. أظهرت دراسة رشاحة مزرعة الفطر قد *A. flavus في الصنف سافيتيري (CR 210-109)*. انخفضت نسبة إنبات البذور، وطول الفروع الجذرية ووزن الفروع الجذرية البادرات بشدة عند غمس البذور في الرشاحة الفطرية المخففة وغير المخففة (30% و25%) وفي حالة الرشاحة غير المخففة كان معدل إنبات البذور وموت البادرات بشدة عند غمس البذور في الرشاحة الفطرية المخففة وغير المحاملة، على التوالي.. كان الوزن المتبقي للبذور أخفض (9.0% و35.0%) وفي حالة الرشاحة غير المخففة كان معدل إنبات البذور وموت البادرات بشدة عند غمس البذور في الرشاحة الفطرية المخففة وغير المعاملة، على التوالي.. كان الوزن المتبقي للبذور أخفض (9.0%) في الشاهد غير المامل وأعلى في الرشاحة عنه (520% لمعاملة الشاهد غير المعاملة، على التوالي.. كان الوزن المتبقي للبذور أخفض (52.0%) في الشاهد غير المعامل وأعلى في الرشاحة الفرية المتائج إلى أن الرشاحة المحتوية على الأفلاتوكسين تداخلت مع العمليات الفيزيولوجية والتي منعت البادرة المتطورة من استخدام السويداء

كلمات مفتاحية، أفلاتوكسين، سافيتري، إنبات، Aspergillus flavus، الرز.

References

- 1. **Deepavali, D.S. and K.W. Nilima.** 2012. Effect of aflatoxin on germination and seedling growth. Archives of Applied Science Research, 4: 2441-2446.
- 2. Bars, L.J. and L.P. Bars. 1992. Fungal contamination of aromatic herbs, aflatoxinogenesis and residues in infusions. Microbiologie Aliments Nutrition, 10: 267-271.
- 3. Islam, M.S., H. Rahman, Z. Pervez, M.R. Mahmud and A. Alam. 2012. Studies on seed borne fungi in rice cultivars grown in non saline tidal zones of patuakhali and their effect on seed germination. Bangladesh Research Publications Journal, 6: 286-290.
- 4. Mangala, U.N., K.R.N. Reddy, C.S. Reddy and K. Muralidharan. 2007. Impact of *Aspergillus flavus* on rice seedling growth and aflatoxin B1 production. Indian Journal of Plant Protection, 35: 76-80.
- 5. Bhat, M.Y. and M. Fazal. 2011. Effect of *Aspergillus flavus* metabolites on wheat seed germination and seedlings growth. Arab Journal of Plant Protection, 29: 139-140.
- Kakde, R.B. and A.M. Chavan. 2010. Determination of toxicity of some fungal metabolites on seed germination and pigment leaching. Journal of Ecobiotechnology, 2/6: 46-55.
- Alpsoy, L. 2010. Inhibitory effect of essential oil on aflatoxin activities. African Journal of Biotechnology, 9: 2474-2481.
- 8. Sweets, L.E. and J.A. Wrather. 2009. Aflatoxin in Corn. University of Missouri. Fisher Delta Research

Received: January 21, 2104; Accepted: June 17, 2014

Center, Agricultural Experiment Station, College of Agriculture, Food and Natural Resources: (http://aes.missouri.edu/delta/croppest/aflacorn.stm).

- 9. Hedayati, M.T., A.C. Pasqualotto, P.A. Warn, P. Bowyer and D.W. Denning. 2007. Aspergillus flavus: human pathogen, allergen and mycotoxin producer. Microbiology, 153: 1677-1692.
- Reddy, C.S., K.R.N. Reddy, R.N. Kumar, G.S. Laha and K. Muralidharan. 2004. Exploration of aflatoxin contamination and its management in rice. Journal of Mycology and Plant Pathology, 34: 816-820.
- **11.** Kumar, V., M.S. Basu and T.P. Rajendran. 2008. Mycotoxin research and mycoflora in some commercially important agricultural commodities. Crop Protection, 27: 891-905.
- 12. Abbas, H.K., W.T. Shier, B.W. Horn and M.A. Weaver. 2004. Cultural methods for aflatoxin detection. Journal of Toxicology. Toxin Reviews, 23: 295-315.
- 13. Janardhan, A., D. Subramanyam, A.P. Kumar, M.R. Pradeep and G. Narasimha. 2011. Aflatoxin impacts on germinating seeds. Annals of Biological Research, 2: 180-188.
- 14. Ahammed, S.K., K. Gopal, M. Munikrishnaiah and D. Subramanyam. 2008. Effect of aflatoxin on shoot and root growth of soybean seedlings. Legume Research, 31: 152-154.

تاريخ الاستلام: 2014/1/21؛ تاريخ الموافقة على النشر: 2014/6/17